

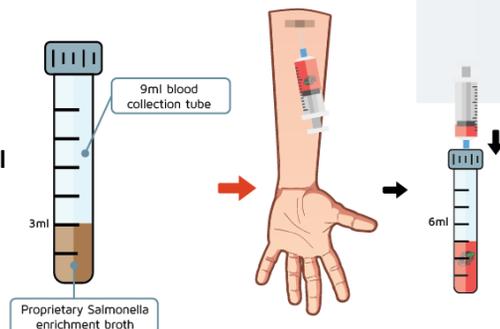
FTIyo Enteric fever

Quick Guide

This product should be used in accordance with Good Laboratory Practice. Do not mix reagents from different lots. Do not use the product after its expiration date.

Step 1: blood collection

- Take a blood collection tube containing the *Salmonella* enrichment broth from the packaging (FTIyo-35-SEB), stored at 4-8°C
- Collect 3 ml of blood from the patient using a syringe
- Pierce the collection tube cap with the needle of the syringe in the vertical position
- Draw the blood in the tube until the liquid reaches the “Max draw volume” black line of the tube label (6 ml)
- Remove the needle from the tube and discard the syringe and the needle

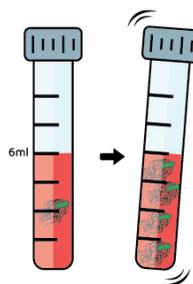


Step 2: sample enrichment

- Set a shaking incubator at 37°C
- Place the blood collection tube filled with the blood sample in the incubator
- Incubate for 5 h* at 180-200 rpm
- Take out the sample from the incubator and process it for nucleic-acid extraction**

* 5 h is the minimum incubation time but it can be increased if necessary

** Samples can be stored for long term storage at -20°C/-80°C before extraction if necessary



Step 3: nucleic-acid extraction

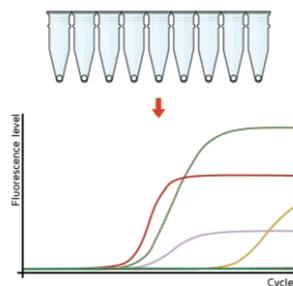
To avoid contamination always handle the enriched sample with gloves and behind a protective shield

- Thaw the negative control (NC, white cap) and the internal control (IC, dark blue cap) found in the packaging (FTIyo-35-32/64-L/-H)
 - Process the NC in parallel to the patient sample
 - Homogenize the enriched blood sample by inverting 5-10 times the blood collection tube
 - Uncap the blood collection tube in a safety cabinet
 - Collect 200 µl of the sample in a separate tube
 - Proceed to nucleic-acid extraction using either the NucliSENS® easyMag® (bioMérieux) specific B protocol or using the QIAamp® DNA blood Mini kit (Qiagen).*
 - Eluate the sample in 55 µl of elution buffer
- *Add 4 µl of IC after the lysis step during nucleic-acid extraction



Step 4: real-time PCR

- Thaw the positive control (PC, red cap)
- Take the necessary amount of PCR tubes containing the lyophilised FTD mastermix
- Add 15 µl of extracted sample, the NC, and the PC to separate lysis mastermix tubes
- Tightly close the tubes with the attached caps
- Slightly vortex the strip and centrifuge briefly
- Place the strip of tubes in the cycler in the vertical position
- Proceed to real-time PCR following FTD cycling profile
- Read Results



FTD cycling profile:
50°C – 15 min – 1x
94°C – 1 min – 1x
94°C – 8 sec – 40x
60°C – 1 min